

# SAMPLING GUIDE

## TISSUE



**Correct sampling procedures and treatment of the tissue samples are important to ensure optimal quality of the analyses. This user manual contains PatoGen's recommendations on how to ensure optimal quality of samples for Real-Time PCR analyses.**

We recommend that the tissue sampling is done by experienced staff. PatoGen will be happy to answer any questions regarding sampling procedures. For statutory analyses samplings shall be performed by an authorized veterinarian, fish health biologist or a designated helper. For instructions on using a helper for parts of the sampling, refer to §6 in FOR 2017-08-29-1318: Regulations on measures to prevent, limit and fight pancreas disease (PD) in aquaculture animals.

### PatoGen Sample Collection Kit:

The kit has been developed as part of PatoGen's quality control system, and consists of pre-labelled tubes with RNAlater™, cooler brick/ gel ice pack, and packaging for transport of the samples to our laboratory. The system is developed to prevent interchange of samples and ensure good quality of the sample tissue that is submitted to PatoGen's laboratory. The kit has a shelf life of six months (expiration date is printed on the side of the kit), and should be stored at room temperature until use.

### Recommended tissue:

| Virus   |                                |   |                     |
|---|--------------------------------|---|---------------------|
| Salmonid alfavirus (SAV)/Pancreas disease virus (PDV)                 | Heart                          | Salmon gill poxvirus (SGPV)                     | Gill                |
| Piscine orthoreovirus (PRV) (HSMI)                                    | Kidney or heart                | Atlantic salmon paramyxovirus (ASPV)            | Gill                |
| Piscine orthoreovirus <i>Oncorhynchus mykiss</i> (PRVom)              | Kidney or heart                | Viral hemorrhagic septicemia virus (VHSV)       | Kidney, brain**     |
| Infectious salmon anaemia virus (ISAV)                                | Heart, kidney, gill*           | Infectious haematopoietic necrosis virus (IHNV) | Kidney              |
| Infectious pancreatic necrosis virus (IPNV)                           | Kidney                         | Nodavirus                                       | Brain               |
| Piscine myocarditis virus (PMCV) (CMS)                                | Heart                          |   |                     |
|   |                                |   |                     |
| Bacteria  |                                |   |                     |
| <i>Renibacterium salmoninarum</i> (BKD)                               | Kidney                         | <i>Candidatus</i> Piscichlamydia salmonis       | Gill                |
| <i>Aeromonas salmonicida</i> subsp. <i>salmonicida</i> (Furunculosis) | Kidney                         | <i>Francisella</i> sp.                          | Kidney              |
| <i>Yersinia ruckeri</i>   | Kidney                         | <i>Moritella viscosa</i>                        | Kidney***           |
| <i>Flavobacterium psychrophilum</i>                                   | Kidney                         | <i>Piscirickettsia salmonis</i>                 | Kidney              |
| <i>Tenacibaculum maritimum</i>  | Gill, kidney, skin             | <i>Candidatus</i> Branchiomonas cysticola       | Gill                |
|   |                                |   |                     |
| Parasites   |                                |   |                     |
| <i>Parvicapsula pseudobranchicola</i> (Myxosporea)                    | Kidney, gill, pseudobranch**** | <i>Ichthyobodo necator</i> (Costia)             | Gill                |
| <i>Paranucleospora theridion</i> (Microsporidia)                      | Kidney                         | <i>Ichthyobodo salmonis</i> (Costia)            | Gill                |
| <i>Paramoeba perurans</i> (AGD)                                       | Gill                           |   |                     |
|   |                                |   |                     |
| Analyses specific to lumpfish   |                                |   |                     |
| <i>Aeromonas salmonicida</i> spp. - atypical (Atypical furunculosis)  | Kidney                         | <i>Pasteurella</i> sp.                          | Kidney              |
| <i>Nucleospora cyclopteri</i>   | Kidney                         | Lumpsucker virus                                | Liver, kidney       |
|   |                                |   |                     |
| Other services  |                                |   |                     |
| SmoltTimer®   | Gill bow                       | Disease Clarification                           | Heart, gill, kidney |

\*For statutory ISA surveillance: Heart, and in some cases also kidney. ISA free areas (establishing new, reestablishing or expansion of ISA free area): Analysis of both heart and kidney is a requirement. Surveillance for ISAV-HPRO: Gill is recommended \*\*Kidney early phase, brain late phase \*\*\*Can also be done on wounds \*\*\*\*Kidney or gill before disease, pseudobranch after

### Preparation

Before sampling and submission, you should have access to:

- Sterile tweezers and scalpels
- Chlorine (10% mix), optionally gas flare and ethanol (70%), for continuous sterilisation of tweezers and scalpel shaft
- PatoGen sample collection kit
- Wipers
- Stationery
- Stable and clean surface



# SAMPLING

## The tissue samples should not get in contact with the interior abdomen of the fish!

Research carried out by PatoGen indicates a certain risk that residual of vaccines can be detected in fish vaccinated against PDV, IPNV, Yersinia or Flavobacter. The risk decreases in time, and is minimized by avoiding direct contact with the vaccine in the internal abdomen. IPNV vaccinated fish are distinguished from infected fish based on which virus variant that are detected, while for other agents there are not yet good methods to distinguish vaccinated and infected fish. Kidney samples should be sampled via the neck for these specific analyses.

### Sterile technique

Use sterile techniques when collecting the samples to avoid contamination between the different samples taken from different fish. We recommend to use a new scalpel for each fish. When reusing equipment we recommend to clean it with wipers to remove organic residuals, before bathing it in 10 % chlorine, optionally you can bathe the equipment in 70% ethanol and burn off. Tip: Use the inside of the scalpel package to trim the tissue.

### Sampling of tissue

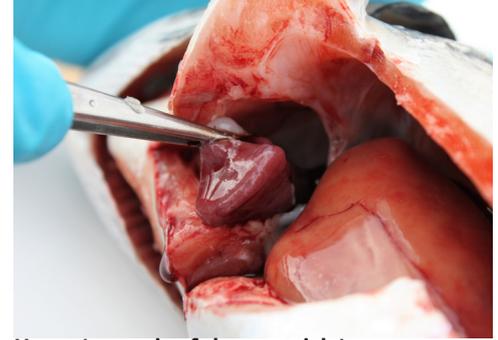
Open the fish with the scalpel so that the relevant organ becomes available. Take the sample, on small fish you can take a larger part of the organs. In case of sampling of gill in addition to other tissues, we recommend taking the gill first (remember to change or secure that the equipment is properly disinfected before you open the fish after sampling of the gill).



Gill



Kidney (head kidney, front part)



Heart (summit of the ventricle)



Trimming of the tissue on the scalpel package



Make sure the tissues are covered by RNAlater



Submission of test tubes in a plastic bag

### Trimming of tissue

Place the tissue on the inside of the scalpel package and cut it into two pieces of about 2x2x2 mm (like the head of a match). Make sure that the tissue samples are not too large as it may cause poor preservation. It is important that you take two samples from each organ being analysed (A and B sample).

### Transfer of tissue to test tubes

The samples from one fish should be placed on the same test tube, and a maximum of four samples can be placed on each tube. If more than one tissue are being sampled (3,5 etc.), we recommend that the gill is placed on a separate tube. Put gill on the first tube and for example heart and kidney on the next. If more than one tissue are being sampled (2, 6 etc.), the gill can be placed on the same tube as an other tissue. Make sure the tissue is placed in the liquid in the tube, and is not stuck on the edge without being covered in liquid.

### Registration of information

Register the sampling in Patolink or note details about the samples on the requisition form following the kit. Contact us if you have any questions regarding access to or use of Patolink. All samplings registered in Patolink gets one day shorter delivery time - 4 working days as opposed to normally 5 working days.

### Storage of the samples before shipment

When the test tubes have been used they should be put on ice/cooler continuously. When doing the sampling over multiple days used test tubes can be stored in a refrigerator for a week before shipment. For longer storage it is recommended that the test tubes are frozen, after having been stored in a refrigerator for 24 hours.

### Shipment

If the whole kit is not used, the test tubes can be removed and placed in a plastic bag. The rest of the kit can then be used in the next sampling. The samples should be sent to PatoGen with a cooling brick/ice pack. Send the package by express shipment to the address below. Send the tracking number to [sample@patogen.no](mailto:sample@patogen.no) or +47 957 07 910.